Murex HIV Ag/Ab Combination

Enzyme immunoassay for improved detection of seroconversion to human immunodeficiency virus types 1 (HIV-1, HIV-1 group O) and detection of anti-HIV-2 antibodies.

Key to symbols used:

- Lot:
- Use by:
- Legal:
- Manufacturer:
- Caution, see instructions for use (potentially infectious):
- Store at 2°C to 8°C:
- For in vitro diagnostic use:
- Consult instructions for use:
- Catalogue number:

See Reagents section for a full explanation of symbols used in component naming.
MUREX HIV Ag/Ab COMBINATION

INTENDED USE
Enzyme immunoassay for improved detection of seroconversion to human immunodeficiency virus types 1 (HIV-1, HIV-1 group O) and detection of anti-HIV-2 antibodies.

SUMMARY AND EXPLANATION OF THE TEST
Two types of human immunodeficiency virus, HIV-1 and HIV-2, have been described and implicated as causative of the Acquired Immunodeficiency Syndrome (AIDS). Both are retroviruses which are transmitted by exposure to certain infected body fluids, primarily blood and genital secretions, and by transplacental passage. Infection by HIV-1 has been reported worldwide; HIV-2 infection has been reported as occurring mainly in West Africa and some European countries.

The two types of virus show substantial antigenic cross reactivity in their gag and pol proteins, but the envelope glycoproteins are less cross reactive.

It is necessary for screening purposes to use epitopes from the envelope proteins of both viruses in addition to major cross reacting gag or pol proteins to ensure detection of antibodies against both types of virus at all stages following infection. Variants of HIV-1, classified together as group O, have been identified in samples from Cameroon and Europe. Group O is highly divergent from the originally known subtypes of HIV-1 (together classified as group M). Specific epitopes from the envelope region of this virus can be used to detect antibody to group O in infected individuals; reliance on cross reactions to the known subtypes of HIV is not satisfactory. The earliest specific antibody response following infection by HIV may be of immunoglobulin M (IgM) followed by a response in immunoglobulin G (IgG). Maximum sensitivity for detection of anti-HIV seroconversion is achieved by assays which respond to both IgM and IgG whilst HIV core antigen is typically detectable during a short period prior to antibody seroconversion.

Murex HIV Ag/Ab Combination is designed to detect reactive HIV core antigen in addition to IgG, IgM and IgA to the envelope glycoproteins and the cross reacting pol proteins of HIV-1 and HIV-2. Consequently, potentially infectious samples of serum, EDTA plasma or citrate plasma can be identified.

PRINCIPLE OF THE PROCEDURE
Murex HIV Ag/Ab Combination is based on microwells coated with a synthetic peptide representing an immunodominant region of HIV-1 (O), recombinant protein derived from the envelope proteins of HIV-1 and HIV-2 and an HIV pol protein and monoclonal antibodies raised against p24 of HIV-1. The Conjugate is a mixture of the same antigen epitopes, and different monoclonal antibodies, also raised against p24, all labelled with horseradish peroxidase.

Test specimens and control sera are incubated in the wells and reactive HIV core and/or antibodies to HIV in the sample or control sera bind to the antibodies and/or antigens on the microwell sample and any excess antibodies are then washed away. In a subsequent step, Conjugate is added which in turn binds to any reactive HIV core and/or specific antibody already bound to the reagents on the well. Samples not containing reactive core antigen or specific antibody will not cause the Conjugate to bind to the well.

Unbound Conjugate is washed away and a solution containing 3,3',5,5'-tetramethylbenzidine (TMB) and hydrogen peroxide is added to the wells. Wells with bound Conjugate develop a blue green colour which is converted to an orange colour which may be read at 450nm after the reaction has been stopped with sulphuric acid.

REAGENTS

DESCRIPTION, PREPARATION FOR USE AND RECOMMENDED STORAGE CONDITIONS
See also Warnings and Precautions.

STORAGE CONDITIONS
All components must be stored at 2 to 8°C, unless otherwise stated, under which condition they will retain activity until the expiry date of the kit.

1. Coated Wells
One plate (7G79-01) or five plates (7G79-02) of 96 microwells coated with HIV antigens and monoclonal antibodies.

Place the wells to reach room temperature (18 to 30°C) before removal from the bag.

2. Sample Diluent
One bottle containing 18ml of a green/brown buffer solution, bovine and murine protein, detergent and saponin. Contains 0.05% ProClin® 300 preservative.

3. Conjugate
One bottle (7G79-01) or three bottles (7G79-02) containing 1.1ml of HIV antigens and monoclonal antibodies conjugated to horseradish peroxidase and freeze dried. When reconstituted each bottle is sufficient for up to two plates.

4. Conjugate Diluent
One bottle (7G79-01) or three bottles (7G79-02) containing 22ml of a yellow solution consisting of buffer, bovine protein, saponin and detergent, sufficient to reconstitute one bottle of Conjugate. Contains 0.1% ProClin® 300 preservative.

Reconstitution of Conjugate
Tap the bottle of Conjugate gently on the bench to remove any material adhering to the rubber stopper. Pour the whole contents of the bottle of conjugate diluent into the bottle of conjugate, recap the latter and mix by gentle inversion. Allow to rehydrate for at least 30 minutes with occasional swirling. The reconstituted conjugate will be red in colour. Reconstituted conjugates may be returned to and pooled in the plastic conjugate diluent bottles if required.

After reconstitution the Conjugate may be stored at 2 to 8°C for up to four weeks.
5. Anti-HIV-1 Positive Control
One bottle containing 1.7ml of inactivated human serum in a buffer containing bovine protein. Contains 0.05% Bronidox® preservative.

6. Anti-HIV-2 Positive Control
One bottle containing 1.7ml of inactivated human serum in a buffer containing bovine protein. Contains 0.05% Bronidox® preservative.

7. HIV-1 p24 Positive Control
One bottle containing 1.7ml of p24 (recombinant antigen) in a buffer containing bovine protein. Contains 0.05% Bronidox® preservative.

8. Negative Control
Two bottles containing 2.5ml of normal human serum. Contains 0.05% Bronidox® preservative.

9. Substrate Diluent
One bottle containing 35ml of a colourless solution of tri-sodium citrate and hydrogen peroxide.

10. Substrate Concentrate
One bottle containing 35ml of 3,3',5,5'-tetramethylbenzidine (TMB) and stabilisers in an orange solution. Substrate Solution
To prepare the Substrate Solution add a volume of colourless Substrate Diluent to an equal volume of orange Substrate Concentrate in a clean glass or plastic vessel.

It is important that this order of addition be followed and that any pipettes and glassware used to prepare Substrate Solution are clean. Alternatively, the Substrate Solution may be made by pouring the entire contents of the bottle of Substrate Diluent into the bottle of Substrate Concentrate. One bottle of Substrate Solution provides sufficient reagent for at least five plates – see Table 1:

### Table 1
Volume of Substrate Concentrate and Substrate Diluent Required

<table>
<thead>
<tr>
<th>Number of Wells</th>
<th>Number of Plates</th>
</tr>
</thead>
<tbody>
<tr>
<td>8</td>
<td>1</td>
</tr>
<tr>
<td>16</td>
<td>2</td>
</tr>
<tr>
<td>24</td>
<td>3</td>
</tr>
<tr>
<td>32</td>
<td>4</td>
</tr>
<tr>
<td>40</td>
<td>1</td>
</tr>
<tr>
<td>48</td>
<td>2</td>
</tr>
<tr>
<td>56</td>
<td>3</td>
</tr>
<tr>
<td>64</td>
<td>4</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Substrate Concentrate (ml)</th>
<th>0.5</th>
<th>1.0</th>
<th>2.0</th>
<th>2.5</th>
<th>2.5</th>
<th>3.0</th>
<th>3.5</th>
<th>4.0</th>
<th>4.5</th>
<th>5.0</th>
<th>6.0</th>
<th>12.0</th>
<th>18.0</th>
<th>22.0</th>
</tr>
</thead>
<tbody>
<tr>
<td>Substrate Diluent (ml)</td>
<td>0.5</td>
<td>1.0</td>
<td>2.0</td>
<td>2.5</td>
<td>3.0</td>
<td>3.5</td>
<td>4.0</td>
<td>4.5</td>
<td>5.0</td>
<td>6.0</td>
<td>12.0</td>
<td>18.0</td>
<td>22.0</td>
<td></td>
</tr>
</tbody>
</table>

Additional reagent may be required for use with automated systems. Keep away from sunlight. The Substrate Solution should be pale yellow; if it is green before being used it should be discarded and fresh Substrate Solution prepared.

The prepared Substrate Solution from this kit may be used interchangeably with that from all other Murex kits which use orange coloured Substrate Concentrate. Ensure that the Substrate Solution is prepared from the Substrate Diluent and Substrate Concentrate provided together.

The prepared Substrate Solution is stable refrigerated (2 to 8°C) or at 15 to 25°C for up to two days but it must be discarded if crystals have formed.

11. Wash Fluid
One (7G79-01) or two (7G79-02) bottles containing 125 ml of 20 times working strength Glycine/Borate Wash Fluid. Contains 0.2% Bronidox® preservative.

Add one volume of Wash Fluid Concentrate to 9 volumes of distilled or deionised water to give the required volume or dilute the entire contents of one bottle of Wash Fluid to a final volume of 2500 ml. Crystals may be observed in the Wash Fluid Concentrate but these crystals will dissolve when the Wash Fluid is diluted to working strength. When diluted the Wash Fluid contains 0.01% Bronidox® preservative.

The Wash Fluid from this kit may be used interchangeably with the Glycine/Borate Wash Fluid from any other Murex kit.

Store the working strength Wash Fluid at 18 to 30°C in a closed vessel under which conditions it will retain activity for one month. NOTE: The Wash Fluid may develop a yellow colour on storage. This will have no effect on the performance of the assay providing the Wash Fluid is fully aspirated from the wells.

NOTE: Although the Substrate Solution and Wash Fluid are interchangeable, they must not be used beyond the expiry date printed on the component labels.
WARNINGS AND PRECAUTIONS

**IVD**
The reagents are for in vitro diagnostic use only.
For professional use only.
Please refer to the manufacturer’s safety data sheet and the product labeling for information on potentially hazardous components.
Low levels of formaldehyde may be observed in the kit controls and product performance is not affected by this. This is a product of certain serum batches used to manufacture the controls.

HEALTH AND SAFETY INFORMATION

CAUTION: This kit contains components of human origin.
The human sera used for manufacture have been screened and found reactive or non-reactive for analytes as shown in Table 2 below:

<table>
<thead>
<tr>
<th>Table 2</th>
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</thead>
<tbody>
<tr>
<td>Component</td>
</tr>
<tr>
<td>Negative Control</td>
</tr>
<tr>
<td>Positive Control 1</td>
</tr>
<tr>
<td>Positive Control 2</td>
</tr>
</tbody>
</table>

All reactive serum used has been inactivated prior to use in reagent preparation. However, all material of human origin should be considered as potentially infectious and it is recommended that this kit and test specimens be handled using established good laboratory practice.

The Sample Diluent contains 0.05% ProClin® 300 and Conjugate Diluent contains 0.1% ProClin® 300 which is classified per applicable European Economic Community (EEC) Directives as irritant (Xi). The following are the appropriate Risk (R) and Safety (S) phrases.

<table>
<thead>
<tr>
<th>Xi</th>
<th>C (C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>R35</td>
<td>Causes severe burns.</td>
</tr>
<tr>
<td>S26</td>
<td>In case of contact with eyes, rinse immediately with plenty of water and seek medical advice.</td>
</tr>
<tr>
<td>S35</td>
<td>This material and its container must be disposed of in a safe way.</td>
</tr>
<tr>
<td>S36/37/39</td>
<td>Wear suitable protective clothing, gloves and/or face protection.</td>
</tr>
<tr>
<td>S46</td>
<td>In case of accident or if you feel unwell, seek medical advice immediately (show label where possible).</td>
</tr>
</tbody>
</table>

Information for European customers: For product not classified as dangerous per European Directive 1999/45/EC - Safety Data Sheet available for professional user on request.

1. Potentially contaminated materials should be disposed of safely according to local requirements.
2. Spillage of potentially infectious material should be removed immediately with absorbent paper tissue and the contaminated area swabbed with, for example, 1% sodium hypochlorite before work is continued. Sodium hypochlorite should not be used on acid-containing spills unless the spill area is first wiped dry.

Materials used to clean spills, including gloves, should be disposed of as potentially biohazardous waste. Do not autoclave materials containing sodium hypochlorite.

3. Neutralised acids and other liquid waste should be decontaminated by adding a sufficient volume of sodium hypochlorite to obtain a final concentration of at least 1.0%. A 30 minute exposure to 1.0% sodium hypochlorite may be necessary to ensure effective decontamination.
4. Do not pipette by mouth. Wear disposable gloves and eye protection while handling specimens and performing the assay. Wash hands thoroughly when finished.
5. If any of the reagents come into contact with the skin or eyes wash the area extensively with water.
6. Sulphuric acid required for the Stop Solution and hydrochloric acid used for washing glassware are corrosive and should be handled with appropriate care. If either come into contact with the skin or eyes, wash thoroughly with water.

**ANALYTICAL PRECAUTIONS**

1. Do not use the reagents beyond the stated expiry date. Microbiological contamination of reagents must be avoided as this may reduce the life of the product and cause erroneous results.
2. Do not modify the Test Procedure or substitute reagents from other manufacturers or other lots unless the reagent is stipulated as interchangeable. Do not reduce any of the recommended incubation times.
3. Allow all reagents and samples to come to 18 to 30°C before use. Immediately after use return reagents to the recommended storage temperature.
4. Any glassware to be used with the reagents should be thoroughly washed with 2M hydrochloric acid and then rinsed with distilled water or high quality deionised water.
5. Avoid the use of self-defrosting freezers for the storage of reagents and samples.
6. Do not expose reagents to strong light or hepatochrome filters during storage or during incubation steps.
7. Do not allow wells to become dry during the assay procedure.
8. Do not cross-contaminate reagents. Dedicated a pipette for use with the Substrate Solution of Murex assays. A pipette should also be dedicated for use with the Conjugate.
9. The Sample Diluent in this assay has the potential to cause false positive results in anti hepatitis B surface antigen (anti-HBs) assays if reagent cross contamination occurs.

If running this assay in conjunction with the Murex anti HBs assay (2K95) on the Tecan Genesis RMP fixed tip instrument ensure that the protocol includes a sodium hydroxide decontamination step after the sample dispense.

If using another manufacturers anti-HBs assay, or any instrument not supported by Abbott, ensure that the possibility of cross contamination is excluded during the validation process.
10. Do not touch or splash the rim of the well with Conjugate. Do not blow out from micropipettes; reverse pipetting is recommended whenever possible.
11. Ensure that the bottom of the plate is clean and dry and that no bubbles are present on the surface of the liquid before reading the plate.
12. Do not contaminate microwells with dust from disposable gloves.
13. When using fully automated microplate processors it is not necessary to use plate lids and tap dry the wells.
14. Do not allow system fluids from automated microplate processors to contaminate the samples or reagents.
15. Ensure the assay is run within the temperature limits defined in the assay protocol.
16. Do not use CO2 incubators.
17. Do not store the Stop Solution in a shallow dish or return it to a stock bottle after use.
18. The possibility of cross contamination between assays needs to be excluded when validating assay protocols on instrumentation.

SPECIMEN COLLECTION, TRANSPORT AND STORAGE

SPECIMEN COLLECTION
Serum, EDTA plasma or citrate plasma samples may be used. Ensure that the serum samples are fully clotted. Remove any visible particulate matter from the sample by centrifugation.

SPECIMEN TRANSPORT AND STORAGE
Store samples at 2 to 8°C. Samples not required for assay within 72 hours should be removed from the clot or cell pellet and stored frozen (-15°C or colder). Avoid multiple freeze-thaw cycles. After thawing ensure samples are thoroughly mixed before testing.

PROCEDURE

MATERIALS REQUIRED BUT NOT PROVIDED
1. Stop Solution (0.5M to 2M Sulphuric Acid), e.g. add between 3.0ml (for 0.5M) and 11ml (for 2.0M) of analytical grade concentrated sulphuric acid (98%) to about 80 ml of distilled or deionised water and then make up to 100ml with more water. Alternatively, the following reagent can be used: 2M Sulphuric Acid (code 7212-05).
2. Freshly distilled or high quality deionised water is required for dilution of Wash Fluid, for preparation of the Stop Solution and for use in conjunction with automated washers.
3. Micropipettes and Multichannel micropipettes of appropriate volume.
4. Incubator capable of maintaining the temperature limits defined in the assay protocol.
5. Moulded Heating Block (Code SF09-02). For use in laboratory incubators. The moulded heating block should ideally be kept in the incubator used. If this is not possible it must be placed in the incubator at least four hours before beginning the assay.
6. Instrumentation
   a) Automated microplate stripwasher.
   b) Microplate reader.
   c) Fully automated microplate processor.
   All instruments must be validated before use. Please contact your representative for details of systems, software protocols for instrumentation and analyser procedures.
8. Sodium hypochlorite. (Refer to Health and Safety Information)
9. Sodium hydroxide solution. (Refer to Analytical Precautions)

TEST PROCEDURE
Please read Analytical Precautions carefully before performing the test.

Addition of the various components of the assay to the wells may be confirmed visually by examining the plate for the following colours:

Sample Diluent is green/brown in colour. On addition of Sample or Control the colour will change to blue/green. The change in colour will vary from sample to sample but some change should always be visible. The addition of sample or control may be confirmed using a microplate reader at 570nm or 620nm with a reference of 690nm.

Conjugate is red in colour. The addition of Conjugate may be confirmed using a microplate reader at 490nm with a reference of 620nm.

Substrate Solution is initially pale yellow with any reactive wells becoming blue green. On addition of Stop Solution the blue green colour of the reagents will change to orange, whilst the negatives will change to pink. The addition of Substrate Solution may be confirmed using a microplate reader at 450nm (no reference).

<table>
<thead>
<tr>
<th>SEMI AUTOMATED PROCESSING</th>
</tr>
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<tbody>
<tr>
<td><strong>Step 1</strong></td>
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<tr>
<td><strong>Step 2</strong></td>
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<td><strong>Step 3</strong></td>
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<td><strong>Step 4</strong></td>
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<td><strong>Step 5</strong></td>
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<td><strong>Step 6</strong></td>
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<tr>
<td><strong>Step 7</strong></td>
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<tr>
<td><strong>Step 8</strong></td>
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<tr>
<td><strong>Step 9</strong></td>
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<tr>
<td><strong>Step 10</strong></td>
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<tr>
<td><strong>Step 11</strong></td>
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<tr>
<td><strong>Step 12</strong></td>
</tr>
<tr>
<td><strong>Step 13</strong></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th><strong>WASH PROCEDURES</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Step 1</strong></td>
</tr>
<tr>
<td><strong>Step 2</strong></td>
</tr>
</tbody>
</table>

NOTE: Do not allow the wells to become dry during the assay procedure. Washers must be rinsed with distilled or deionised water at the end of the test to avoid blockage and corrosion.
FULLY AUTOMATED PROCESSORS
Validated protocols for a range of automated instruments are available, contact your representative for details. For instrumentation without established validated protocols, the following guidelines are recommended:

1. Do not programme times shorter than specified in the procedure.
2. For each incubation at 37°C, programmed times may be increased by up to 5 minutes.
3. Wells containing Sample Diluent may be left for up to 60 minutes at 18-36°C prior to the addition of Sample and for up to 60 minutes after the addition of samples or Controls before starting step 5 in the assay protocol.
4. Ensure all Analytical Precautions are followed.

Protocols written following these guidelines must be fully validated prior to use according to local procedures.

RESULTS
CALCULATION OF RESULTS
Each plate must be considered separately when calculating and interpreting results of the assay. Approved software may be used for calculation and interpretation of results.

Negative Control
Calculate the mean absorbance of the Negative Controls. Example:

Well 1 = 0.084, Well 2 = 0.086, Well 3 = 0.070
Mean Negative Control = (0.084 + 0.086 + 0.070) / 3 = 0.080

If one of the Negative Control Wells has an absorbance more than 0.15 O.D. above the mean of all three, discard that value and calculate the new Negative Control mean from two remaining replicates.

Cut-off value
Calculate the Cut-off value by adding 0.150 to the mean of the Negative Control replicates (see above).

Mean Negative Control = 0.080
Cut-off value = 0.080 + 0.150 = 0.230

QUALITY CONTROL
Results of an assay are valid if the following criteria for the Controls are met:

Negative Control
The mean absorbance is less than 0.15.

Positive Controls
The absorbance of each of the Positive Controls is more than 0.8 above the mean absorbance of the Negative Control.

Assays which do not meet these criteria should be repeated. In the unlikely event of the results repeatedly failing to meet either the Quality Control criteria or the expected performance of the test, please contact your representative.

INTERPRETATION OF RESULTS
Non-reactive Results
Samples giving an absorbance less than the Cut-off value are considered negative in the assay.

Reactive Results
Samples giving an absorbance equal to or greater than the Cut-off value are considered initially reactive in the assay (see Limitations of the Procedure).

Unless local procedures state otherwise, such samples must be retested in duplicate using the original source. Samples that are reactive in at least one of the duplicate retests are considered repeatedly reactive in Murex HIV Ag/Ab Combination and are presumed to contain reactive HIV core antigen and/or antibodies to HIV-1 or HIV-2. Such samples must be further investigated and the results of this assay considered with any other clinical and/or assay information. Samples that are non-reactive in both wells on retest are considered non-reactive for HIV core antigen and HIV antibodies.

No sample addition
Absorbance values significantly higher than the Negative Control may be obtained in wells where the sample has been omitted but all the reagents have been added.

SPECIFIC PERFORMANCE CHARACTERISTICS
The performance of Murex HIV Ag/Ab Combination has been determined by testing samples from random blood donors, patients with AIDS diagnosed according to CDC criteria, patients with AIDS Related Complex (ARC), other patients with known antibody to HIV-1 (including group O), patients with confirmed HIV-2 infection and patients at risk of HIV infection or in other clinical categories. In addition, its performance on commercially available serocconversion panels has been evaluated.

Diagnostic Sensitivity
A total of 497 specimens from patients with confirmed HIV-1 infection were tested and found to be reactive with Murex HIV Ag/Ab Combination. The specimens were taken from patients at various stages of HIV infection and included 24 specimens from patients with HIV-1 subtype O infection and a further 139 specimens from patients infected with HIV-1 subtypes other than subtype B.

In addition a total of 100 specimens from patients with confirmed HIV-2 infection were also tested with Murex HIV Ag/Ab Combination and found to be reactive.

The diagnostic sensitivity of Murex HIV Ag/Ab Combination on this population of specimens is therefore estimated to be 100% (597/597) with a lower 95% confidence limit of 99.38% (593/597) by the binomial distribution.

A total of 26 commercial HIV-1 serocconversion panels were tested with Murex HIV Ag/Ab Combination. Using the presence of both core (p24) and an envelope (gp120/160) band on Western blot as the reference criteria, Murex HIV Ag/Ab Combination detected antibody to HIV earlier or in the same sample as Western blot in all of the panels.

Diagnostic Specificity
In a study where specimens from a European blood donor population were tested. A total of 3,950 routine donor plasma specimens were screened with Murex HIV Ag/Ab Combination at three European blood transfusion centres. The results are summarised in Table 3. In the study, 99.78% (9289/9329) of specimens were non-reactive and 0.22% (20/9329) were reactive. One of the specimens was weakly positive with the Murex HIV Antigen Mab (8E77).

The diagnostic specificity of the Murex HIV Ag/Ab Combination on presumed negative European blood donors is estimated to be 99.78% (9289/9329) with 95% confidence limits of 99.67% (9258/9289) to 99.86% (9276/9289) by the binomial distribution.

A total of 267 specimens from patients with conditions unlinked to HIV infection were also tested with Murex HIV Ag/Ab Combination. These included specimens from pregnant women and patients suffering with autoimmune disease and other acute viral infections. A total of five specimens were reactive with Murex HIV Ag(AB); four were reactive with two other commercially available screening assays. In Western blot studies four produced indeterminate results and one was negative.

In addition, 38 lipaemic, icteric and haemolysed specimens were also tested and found to be non-reactive.

The overall diagnostic specificity of Murex HIV Ag/Ab Combination on confirmed negative specimens during this performance evaluation is estimated to be 99.78% (9289/9300) with 95% confidence limits of 99.67% (9258/9290) to 99.87% (9276/9290) by the binomial distribution.

*Representative performance data are shown. Results obtained at individual laboratories and with different populations may vary.

Assay Reproducibility
The reproducibility of Murex HIV Ag/Ab Combination was assessed by testing two of the assay controls and four quality assurance panel members as ten replicates on four separate occasions. The results from the testing are summarised in Table 4.
Table 3
Reactivity of Murex HIV Ag/Ab Combination with presumed negative specimens from routine European blood donors.

<table>
<thead>
<tr>
<th>Centre</th>
<th>Number of presumed negative specimens tested</th>
<th>Number of repeatedly reactive specimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>3095</td>
<td>6 (0.19%)</td>
</tr>
<tr>
<td>B</td>
<td>2803</td>
<td>9 (0.32%)</td>
</tr>
<tr>
<td>C</td>
<td>3392</td>
<td>6 (0.18%)</td>
</tr>
<tr>
<td>TOTAL</td>
<td>9290</td>
<td>21 (0.23%)</td>
</tr>
</tbody>
</table>

* Includes one specimen which was weakly positive in Murex HIV Antigen Mab (8E77).

Table 4
Murex HIV Ag/Ab Combination - Assay Reproducibility

<table>
<thead>
<tr>
<th>Specimen</th>
<th>Number of Assays</th>
<th>Number of Replicates</th>
<th>Mean Absorbance/Cut-off ratio</th>
<th>Intra-assay % CV</th>
<th>Inter-assay % CV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative Control</td>
<td>4</td>
<td>10</td>
<td>0.265</td>
<td>8.7</td>
<td>11.3</td>
</tr>
<tr>
<td>HIV-1 Positive Control</td>
<td>4</td>
<td>10</td>
<td>0.287</td>
<td>4.3</td>
<td>4.7</td>
</tr>
<tr>
<td>QA Q1</td>
<td>4</td>
<td>10</td>
<td>3.672</td>
<td>4.6</td>
<td>7.3</td>
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<tr>
<td>QA Q2</td>
<td>4</td>
<td>10</td>
<td>4.696</td>
<td>5.6</td>
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<tr>
<td>QA Q3</td>
<td>4</td>
<td>10</td>
<td>3.006</td>
<td>3.9</td>
<td>8.1</td>
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<tr>
<td>QA Q4</td>
<td>4</td>
<td>10</td>
<td>1.863</td>
<td>5.8</td>
<td>9.2</td>
</tr>
</tbody>
</table>

Sensitivity on AFSSAPS HIV Ag Standard

Sensitivity of Murex HIV Ag/Ab Combination on the AFSSAPS HIV Ag standard was determined at three testing centres.

Table 5
Sensitivity on AFSSAPS HIV Ag standard

<table>
<thead>
<tr>
<th>Centre</th>
<th>Sensitivity HIV Ag pg/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
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</tr>
<tr>
<td>2</td>
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</tr>
<tr>
<td>3</td>
<td>25</td>
</tr>
<tr>
<td>Mean</td>
<td>28</td>
</tr>
</tbody>
</table>

The data shown in Table 5 was obtained during this testing but may not be exactly reproducible on other testing occasions.

LIMITATIONS OF THE PROCEDURE
1. The Test Procedure and Interpretation of Results must be followed.
2. This test has only been evaluated for use with individual (unpooled) serum, EDTA plasma or citrate plasma samples.
3. A negative result with an antigen/antibody detection test does not preclude the possibility of infection with HIV.
4. A positive result with Murex HIV Ag/Ab Combination should be confirmed by at least one other test.
5. This test may give false results with some samples containing HIV-2 antibodies.
6. Non-repeatable reactive results may be obtained with any EIA procedure.

The most common sources of error are:
- Improper delivery of Sample, Conjugate or Substrate into the wells.
- Contamination of Substrate with Conjugate.
- Contamination with conjugates from other assays.
- Blocked or partially blocked washer probes.
- Insufficient aspiration leaving a small volume of Wash Fluid in the wells.
- Failure to ensure that the bottom surface of the wells is clean and dry, and that no air bubbles are present on the surface of the liquid in the wells before a plate is read.
- Failure to read at the correct wavelength (450 nm) or use of an incorrect reference wavelength (not 620 nm to 690 nm).
- Use of highly haemolysed samples, incompletely clotted sera, plasma samples containing fibrin or samples with microbial contamination may give rise to erroneous results.
- This test has not been evaluated for use with samples from cadavers.

BIBLIOGRAPHY

Murex® and ProClin® are not trade marks of Abbott

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